Kacey Diagnostics C&S Products and Accessories

| Product Description | Order Number | Unit Measure | MSRP |
|--|-----------------|-----------------|----------|
| MutiChrome 10 pack | 20203 | 10 Pak | \$130.00 |
| MutiChrome Mastitis 10 pack | 20208 | 10 Pak | \$110.00 |
| Dermatophyte Plates 10 pack | 20210 | 10 Pak | \$80.00 |
| Kacey Mini Incubator (6 samples) | 20211 | 1 Ea | \$695.00 |
| Kacey Maxi Incubator (12 samples) Special Order | 20212 | 1 Ea | CALL |
| Culture Swabs 100% Rayon Sterile Individual | 20216 | 100 Pak | \$40.00 |
| Culture Loops 10uL Sterile Individual | 20217 | 100 Pak | \$40.00 |
| Gram Negative Sensi-Ring | 20221 | 10 Pak | \$100.00 |
| Gram Positive Sensi-Ring | 20222 | 10 Pak | \$110.00 |
| Ear Sensi-Ring | 20223 | 10 Pak | \$130.00 |
| UTI Sensi-Ring | 20224 | 10 Pak | \$130.00 |
| Skin & Wound Sensi-Ring | 20226 | 10 Pak | \$120.00 |
| Kacey Maxi Muller Hinton Plate 100mm | 20234 | 10 Pak | \$85.00 |
| Working Solution Tube (0.85%) | 20235 | 25/Box | \$60.00 |
| Quick Reference Guide | 20236 | 50/Box | \$5.00 |
| Quick Reference Overlay Plastic Reader | 20237 | 1 Ea | \$5.00 |
| Quick Reference Inhibition Chart | 20238 | 1 Ea | No Chg |
| Patient Lab Report Form (2 copies) | 20240 | 25 Pak | \$25.00 |
| 4 Test Starter C&S Kit FOUR TEST STARTER KIT CONSISTS OF THE FOLLOWING ITEMS IN THE KIT: Contents 4 Ea. Multichrome Bi-plates 4 Ea. Kacey Maxi-Muller Hinton Plates (TRUE-100mm) 5 Ea. Sterile Loops (10 uL) | 20241 | 1 Kit | \$125.00 |

1 EA. Standard Turbidity Tube

1 Ea. Inhibition Overlay Reader

1 EA. Sensi Ring-EAR, UTI, GRAM (-) & GRAM (+)

Plus Bonus Wound / Skin Sensi Ring

Ea. Quick Reference Chart
Ea. Instruction Manual Insert
Ea. Patient Report Form -2 copy



Can Now Detect MRSP With Sensi-RingTM

Skin/Wound

#20226

V.O.L.T.™ Veterinary Office Laboratory Tests

INSTRUCTION MANUAL



Thank you for purchasing the Kacey Diagnostics "C& S" Culture and Sensitivity products featuring the "MultiChrome"™ Bi-Plate for cultures with Chromogenic technology and the "Sensi-Rings"™, a self contained ring of antibiotic disks.

Cutting edge Bacteriology, that's easy, accurate and in house, at a fraction of the cost of outside labs!

Culture

PRINCIPLE OF THE TEST

If there is the presence of bacteria MultiChrome[™] Bi-Plate will interpret this in the following manner. The gram (+) side of the bi-plate will only be able to detect gram (+) bacteria. Conversely the Gram (-) side of the plate will allow only gram (-) bacteria to grow. MultiChrome will first confirm the presence of bacteria, followed by separation into their respective gram positive and negative families. Ultimately the bacteria will exhibit a distinctive color through the MultiChrome[™] Bi-Plate process.

Example: (SEE LAMINATED COLOR CHART FOR ALL BACTERIA PICTURES)

Gram Positive Bacteria

Enterococcus= small, teal to turquoise colonies Staph saprophyticus= opaque, pink colonies Staph aureus= large, white center with milky white border

Gram Negative Bacteria

E. coli= med size, rose to magenta colonies with darker center Pseudomonas a. =light yellow green, translucent with iridescense Klebsiella p.=large, dark blue or indigo colonies

SPECIMEN COLLECTION AND INNOCULATION OF BI-PLATE

1) Remove the plate from refrigerator & pre-heat in the incubator @37 degrees for 15-20 minutes prior to inoculating the bi-plate.

2) Due to the diversity of bacterial sites it is recommended that you review standard references for the proper collection of the sample at these different collection sites. If these are concerns regarding infectious material, the sample should be immediately tested or protected from excessive heat and cold. It is recommended that if there is a delay in testing, the specimen should be properly stored in a sealed container and placed in the refrigerator until inoculation of the Bi-Plate at a later time.

3) Use a Kacey Sterile inoculating loop (20uL) or a Sterile Rayon Swab and inoculate the sample onto both sides of the bi-plate by utilizing a zigzag streaking motion. (See Quick Reference Card with illustrations that is included with each kit)

3) Re-place clear lid and place bi-plate in the incubator upside down (inverted position) and incubate at 37°C +/- 2 degree C for no less then 24 hours . The plate should be examined after 24 hours, but no later than 48 hours after incubation for colony growth showing both typical morphology and the specific color unique to that organism. Yeast growth may require a longer period of time to grow and therefore may require a total of 48 hours for adequate incubation growth to occur. (If using Kacey Micro Incubator see instructions for complete details)

Note: Each Kacey C&S Kit comes with a "Quick Reference Card" with Test Procedural illustrations.

PRECAUTIONS

*For in vitro diagnostic Veterinary use only.

*Out of date MultiChrome should never be used to perform a test. Check each plate for the expiration date. Each clinic should abide by their local city, county, and state regulations for the proper disposal of biohazard animal waste. Use recognized safety protocols when handling and disposing of cultured MultiChrome media. You are culturing live active bacteria that may be zoonotic. Agents such as leptospirosis, e. coli, s. aureus, etc can be transmitted. Use of gloves, mask or safety shield is recommended. Wash after handling.

Sensitivity

PROCEDURE FOR INOCULATING THE KACEY MAXI-MULLER HINTON PLATE & SENSITIVITY TESTING The Kacey Sensi-Rings can only fit on the Kacey Maxi Muller Hinton Plates to assure proper antibiotic diffusion and virtually no cross over of antibiotic zones. All other Muller Hinton Plates with "standard Size Dimensions" will not work with the Kacey Sensi-Rings and will render erroneous results.

Kacey Sensi-ring \mathbb{M} (SR \mathbb{M}) should be used according to an appropriate standardized susceptibility test method. Various alternative methodologies are available and Sensi-Ring \mathbb{M} is compatible with these.

- 1) Taking a Kacey Sterile Swab or 10uL Loop and carefully dab on to three (3) different places containing the bacteria on the MultiChrome Bi-plate.
- 2) Immediately place swab or loop containing bacteria into the Kacey Working Solution Tube ("WST")
- Mix the swab with a gentle twirling motion while in the WST tube for approximately 3-5 seconds. Use a swab for inoculation of the Kacey Maxi Muller Hinton (MH) plate.
- Using the now diluted swab streak the MH plate using wide broad strokes starting at the 9-3 o'clock position, followed by 11-5 o'clock and finally the1-7 o'clock position and around the periphery of the Kacey Maxi-MH Plate (See Quick Reference Card illustrations) Discard the swab.

5) Remove the SR[™] pouch from the refrigerator and allow to equilibrate to room temperature before opening. Remove SR[™] from the foil pouch with tweezers at the inner tab & place it onto the Maxi MH Plate "**FACE DOWN**", tapping down in non-disk areas. **(See Quick Reference Card illustrations)** Label the specimen to be incubated.

(6).Place the MH inoculated plate into a 37°C +/- 2 degree preheated incubator **upside down**, set timer inoculate and incubate for 24 hours. Remove & read inhibition zones with the Kacey Digital Reader caliper or Zone Inhibitor Reader enclosed.

Interpretation of results

Interpret measured zones of inhibition by reference to the chart published on the "Quick Reference Card" OR ON THE Patient Lab Report Form enclosed with each starter kit which classify test isolate as

Susceptible (S) Intermediate (I) - Resistant (R).

Good Quality Control laboratory procedures are determined by checking for signs of deterioration. If QC is required it must be performed with at least one organism to demonstrate a correct susceptibility pattern. Do not use the product if the reactions with the control organisms are incorrect. The list below illustrates a range of performance control strains which the end user can easily obtain from Kacey by calling 828.685.3569

| Test Organisms | | | |
|------------------------|------------------------|--|--|
| Escherichia coli | Correct susceptibility | | |
| ATCCâ 25922 | pattern* | | |
| Pseudomonas aeruginosa | Correct susceptibility | | |
| ATCCâ 27853 | pattern* | | |
| Staphylococcus aureus | Correct susceptibility | | |
| ATCCâ 25923 | pattern* | | |

Limitations

Any deviation from the prescribed method may produce incorrect results. The latest published version of the method used should be consulted for complete details of test procedures and interpretive results.